Preventive and therapeutic effects of NF-kappaB inhibitor curcumin in rats colitis induced by trinitrobenzene sulfonic acid

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INTRODUCTION

Intestinal mucosa has functions of excretion, absorbability, immunity and barrier. Mucosal barrier function is composed of intestinal epithelial cells, components outside cells and immune system. They can block deleterious matter from entering the body. Mucosal barrier function is preternatural when intestinal inflammation occurs. Pathogenesis of inflammatory bowel disease (IBD) is ambiguous. IBD includes ulcerative colitis (UC) and Crohn’s disease (CD) in the clinic. Both of them are characterized by mucosal inflammation in pathologic histology[1,2]. The common point in mucosal inflammation research has been offered for preventing and treating IBD. It is important for scientists in the digestive domain. Intestinal mucosal inflammation induced by environment, descendiblity and immunity. Immunity is crucial above all. Homeostasis of TH1/TH2 type cytokines is important in intestinal mucosal immunity. Proinflammatory cytokines were significantly enhanced and anti-inflammatory cytokines were decreasing in intestinal inflammation.

Incidence of IBD is high in the western countries. It is increasing in countries of Asia year after year. Treatments for IBD in clinic cause big side effects. Their aims are ambiguous and they are costly. Their curative effects are not satisfying. Filtrating high effects and low toxicity of natural anti-inflammatory medications are desired for the treatment of patients with IBD. Curcumin is a natural plant phenolic active component. Spice curcumin has different pharmacologic effects, including antitumor, antioxidation and anti-inflammatory modulation of immunity functions[3–6]. The pleiotropic effects of curcumin owe to inhibition of NF-κB-inducing kinase and IκB kinase in intestinal epithelial cells[7]. The study was promising to ascertain via molecule mechanism whether NF-κB inhibitor curcumin prevents and cures intestinal mucosal inflammation in IBD. Using rats colitis model induced by TNBS, this research is to ascertain curcumin’s preventive and therapeutic effects of NF-κB inhibitor curcumin in rats colitis induced by trinitrobenzene sulfonic acid.

Abstract

AIM: To ascertain the molecule mechanism of nuclear factor-κB (NF-κB) inhibitor curcumin preventive and therapeutic effects in rats’ colitis induced by trinitrobenzene sulfonic acid (TNBS).

METHODS: Sixty rats with TNBS-induced colitis were treated with 2.0% curcumin in the diet. Thirty positive control rats were treated with 0.5% sulfasalazine (SASP). Thirty negative control rats and thirty model rats were treated with general diet. Changes of body weight together with histological scores were evaluated. Survival rates were also evaluated. Cell nuclear NF-κB activity in colonic mucosa was evaluated by using electrophoretic mobility shift assay. Cytoplasmic IκB protein in colonic mucosa was detected by using Western Blot analysis. Cytokine messenger expression in colonic tissue was assessed by using semiquantitative reverse-transcription polymerase chain reaction.

RESULTS: Treatment with curcumin could prevent and treat both wasting and histopathologic signs of rats with TNBS-induced intestinal inflammation. In accordance with these findings, NF-κB activation in colonic mucosa was suppressed in the curcumin-treated groups. Degradations of cytoplasmic IκB protein in colonic mucosa were blocked by curcumin treatment. Proinflammatory cytokine messenger RNA expression in colonic mucosa was also suppressed.

CONCLUSION: This study shows that NF-κB inhibitor curcumin could prevent and improve experimental colitis in murine model with inflammatory bowel disease (IBD). The findings suggest that NF-κB inhibitor curcumin could be a potential target for the patients with IBD.
MATERIALS AND METHODS

Animals and grouping
In this study, 10-12 wk old and 200-250 g male SPF Wistar rats were used. All rats were bred under clean barrier conditions in Animal Laboratorical Center of First Military Medical University. Rats were randomly divided into 5 groups: negative control group, positive model group, treatment positive control group, curcumin preventive group, curcumin therapeutic group.

Model of colitis and treatment
To induce intestinal inflammation model by portion of 50 mg hapten TNBS (Sigma Company) in 2 mL 50% ethanol. 2 mL mixed liquor was fleetly injected into the intestine of each group of Wistar rats while negative control rats received 2 mL 50% ethanol.

Curcumin (purity, 95%) was purchased from BDH Company in England. For dietary administration, curcumin was mixed with the feed to concentrations 20%. Preventive group rats were administered 20% curcumin starting 3 d before administration of TNBS (group 4, 2.0% CUR-P). Therapeutic group rats were administered 20% curcumin just after administration of TNBS (group 5, 2.0% CUR-A). Sulfasalazine (SASP) was mixed with the feed to a concentration of 0.5%. Treatment positive control rats were treated with 0.5% SASP starting 3 d before administration of TNBS (group 3, 0.5% SASP). Negative control rats were treated with general feed after administration of 50% ethanol (group 1, ETHA). Positive model rats were treated with general feed (group 2, TNBS). Two weeks after administration of TNBS, all the rats were killed.

Histological analysis and grades
For histological analysis, rats’ colons were fixed in 40 g/L formaldehyde, and paraffin-embedded tissue sections were stained with HE (hematoxylin and eosin) using standard techniques. Histological changes were graded: 0, no signs of inflammation; 1, very low level of leukocyte infiltration; 2, low level of leukocyte infiltration; 3, high level of leukocyte infiltration, high vascular density, and thickening of the colon wall; 4, leukocyte transmural infiltration, loss of goblet cells, high vascular density and thickening of the colon wall.

NF-κB activation analysis
Proteins extracts were prepared according to the method of the Nuclear Extract Kit (Active Motif Company). The Nuclear Extract Kit has been developed for the preparation of nuclear, cytoplasmic extracts from rat’s tissue. Protein was stored at 80 °C. Protein concentrations were measured by Bradford assay.

For electrophoretic mobility shift assay to monitor NF-κB activation of intestinal mucosa, 10 μg of nuclear extracts was preincubated- in 10 μL reaction buffer [2 ng poly dI_dC (Sigma Company), 25% glycerol, 1 mol/L NaCl, 10 mmol/L MgCl₂, 10 mmol/L DTT, 0.2 mol/L Tris HCl, 10 mmol/L EDTA] for 15 min at room temperature. As a control measure, 1 μL 200-fold molar excess of cold NF-κB competitor oligonucleotide probe was added during preincubation. After pre-incubation, 25 000-50 000 CPM 32P end-labeled NF-κB oligonucleotide hot probe (5’-AGTTGGGGACTTTCCCAGGC-3’; 3’-CTCTCCCTGAAAGGGTTCCGTTGA-5’; composed by Shanghai Sangon Biological Engineering Company) was added to the reaction mixture and incubated for 30 min. The reaction mixture was loaded onto a 8% polyacrylamide gel containing Tris-borate-ethylenediamine-tetracetic acid buffer and electrophoresis was performed at 130 V for 3 h. Gels were dried and exposed to Kodak X-ray film.

IkB degradations analysis
Using the same technique as herein before narrated for cytoplasmic protein extracts and protein concentrations measure. Using Western blot analysis to monitor degradations of cytoplasmic IkB protein of intestinal mucosa. For each sample, equal amounts (50 μg, 20 μL) of protein lysates were heated to 97 °C for 5 min and loaded onto 12% SDS-PAGE gel. Proteins were electrotransferred to PVDF membranes (Bio-Rad Company) by BIO-RAD TRANS-BLOT SEMI-DRY TRANSFER CELL apparatus, blocked in Blocking Solution (Biocytex) for 1 h at room temperature, and then incubated membrane and 1:500 rabbit polyclonal anti-IκBα primary antibody overnight at 4 °C. (Santa Cruz Biotechnology Company). Incubated membrane with 1:5 000 Peroxidase-Conjugated (HRP-conjugated) secondary antibodies for 1 h at room temperature, and then to detect protein marker in ECL Western blotting detection reagents (Amersham Company). Exposed to Kodak X-ray film are developed.

Cytokine IL1β and IL10 mRNA analysis
Semi-quantitative RT-PCR analysis of IL1β and IL10 mRNA was extracted from total RNA from 50 to 100 mg colon mucosa with TRIzol Reagent (Invitrogen company). The 1 μg RNA was converted to complementary DNA using random hexonucleotides in 10 μL of reverse-transcription reaction solution and then used for polymerase chain reaction, and then using Oligo-dT Primer and MuLV reverse transcriptases (Promega Company). For PCR amplification, one-fourth of the reverse transcription product and 10 mmol/L of each oligonucleotide as primers were used. After one denaturation step at 95 °C for 5 min, 30 cycles of amplification were performed: denaturation at 94 ºC for 30 s, annealing at 47 ºC for 45 s, synthesis at 72 ºC for 1 min, and extension at 72 ºC for 5 min. The primers used were as follows: to check cDNA quality, GAPDH sense: 5’-ACCACAGTCCATGGCATCAC-3’, GAPDH antisense:5’-TCCACACACCCTGGTCTGTA-3’ (452 bp); IL1β sense:5’-CTTCCTCTTTGAGTTGATTTT-3’, IL1β antisense:5’-CTTCCTCTTTGAGTTGATTTT-3’ (325 bp); IL10 sense:5’-GCAAGAACAAACATCCG-3’, IL10 antisense:5’-ACTTTGCCCTCATCCC-3’ (264 bp). One-tenth of the amplified products was run on 1% agarose gels in 1×Tris-borate-ethylenediamine-tetracetic acid buffer and visualized with ethidium bromide.
Statistics analysis
The results of rats’ body weight were expressed as mean±SE. The survival rates comparison was determined by the χ² test, with P<0.05 considered significant. Statistics of histologic scores were determined by Two-Independent Samples Tests, with P<0.05 was considered significant. All data were automatically performed by statistical software.

RESULTS
Effects of curcumin on the survival rate of rats with TNBS-induced colitis
As shown in Figure 1, the survival-rate of positive model rats with TNBS-induced colitis was 60%. The survival rate of negative control rats with 50% ethanol was 100%. In preventive and therapeutic model, the survival rates of rats were 93.33% and 80%. The survival rate of positive control rats treated with 0.5% SASP was 76.67%. Compared to that of rats with colitis, the survival rate of rats treated with medicine was significantly higher. However, the survival rate of 2.0% curcumin preventive mode is particular (against group 2, P<0.02). According to the data, preventive treatment with curcumin is capable of increasing the survival rate of rats with colitis induced by TNBS.

Effects of curcumin on body weight of rat with TNBS-induced colitis
As shown in Figure 2, body weights of positive model rats, with TNBS-induced colitis, had obviously decreased (more than 15% after 2 d); body weights had recovered gradually from d 6 but not fully to the initial weights in 2 wk. Negative control rats receiving 50% ethanol without TNBS had gained weights. However, body weights had increased to 113.3% of the initial weights in 2 wk. In all groups of rats with treatment, body weights recovered from d 2. Body weights were significantly higher than those of untreated positive model rats on d 2-14. Body weights in the group of 50% ethanol were significantly higher than those of other groups (P<0.05). Body weights in the group of 2.0% curcumin and 0.5% SASP-treated rats were significantly lower than that of untreated rats with colitis untreated groups. In this experiment, the histologic scores of rats with TNBS-induced colitis, many dispersive and focal ulcers were detected. Dilipation of tissues epithelium induced inflammatory infiltration. There was a very high level of leukocyte infiltration accompanied with obvious tissues putrescence. In the mucosa of rats of 2.0% curcumin preventive and therapeutic groups, low level of inflammation was detected. Mucosal inflammation of curcumin groups was significantly decreasing than that of untreated groups. In this experiment, the histologic scores of 2.0% curcumin and 0.5% SASP-treated rats were significantly lower than that of untreated rats with colitis (50% ethanol group against TNBS group P<0.01, TNBS group against three medication groups P<0.05).

Effects of curcumin on histologic scores of rats with TNBS-induced colitis
We have characterized the histological features of colitis in rats subjected to TNBS enema by staining with HE (Figures 3A-E). Then we evaluated the effects of curcumin on TNBS-induced colon lesions on histologic scores. The results of histologic scores were shown in Table 1. In the mucosa of rats with TNBS-induced colitis, many dispersive and focal ulcers were detected. Dilipation of tissues epithelium induced inflammatory infiltration. There was a very high level of leukocyte infiltration accompanied with obvious tissues putrescence. In the mucosa of rats of 2.0% curcumin preventive and therapeutic groups, low level of inflammation was detected. Mucosal inflammation of curcumin groups was significantly decreasing than that of untreated groups. In this experiment, the histologic scores of 2.0% curcumin and 0.5% SASP-treated rats were significantly lower than that of untreated rats with colitis (50% ethanol group against TNBS group P<0.01, TNBS group against three medication groups P<0.05).

Effects of curcumin on NF-κB activity of colonic mucosa in rats with TNBS-induced colitis
As shown in Figure 4, we performed electrophoretic mobility shift assay using nuclear extracts of colonic mucosa from each model rat. Unlabeled specific oligonucleotides of 200-fold molar excess inhibited NF-κB mobility shift (Lane 2), indicating that the DNA-protein complex is specific. This land was cold probe control for the other lands. Unlabeled probes of 200-fold molar excess were added to the reaction

![Figure 1](image) Effects of curcumin on the survival rate of rats with TNBS-induced colitis. Group 1: 50% ethanol group. Group 2: TNBS group. Group 3: 0.5% SASP group. Group 4: 2.0% curcumin preventive group. Group 5: 2.0% curcumin therapeutic group.

![Figure 2](image) Effects of curcumin on body weights of rats with TNBS-induced colitis. ■: 50% ethanol group. ●: TNBS group. ◊: 0.5% SASP group. ●: 2.0% curcumin preventive group. ▲: 2.0% curcumin therapeutic group.

| Table 1: Histologic scores of rats with TNBS-induced colitis |
|---|---|---|---|---|---|
| Group | Cases | 0Grade | 1Grade | 2Grade | 3Grade |
| ETHA | 30 | 9* | 17b | 3b | 1b | 0* |
| TNBS | 18 | 0 | 2 | 4 | 5 | 7 |
| 0.5%SASP | 23 | 0* | 6* | 9* | 6* | 2* |
| 2%CUR-P | 28 | 1* | 14* | 9* | 3* | 1* |
| 2%CUR-A | 24 | 0* | 7* | 10* | 5* | 2* |
| Count | 123 | 10 | 46 | 35 | 20 | 12 |

*p<0.05 ; b<0.01 vs TNBS group.

![Image](image)
mixture of lane 3 (Lane 2). The administration of 50% ethanol had not activated NF-κB DNA binding activity of nuclear extracts in the colonic mucosa. The administration of TNBS alone had enhanced NF-κB DNA binding activity (Lanes 3, 4), which was suppressed by treatment with 0.5% SASP (Lane 5) and 2.0% curcumin (Lanes 6, 7).

**Effects of curcumin on cytoplasmic IκB degradations**

Since NF-κB activity is controlled by the steady state level of IκB, we further investigated the effect of curcumin on IκB degradation in rat’s colonic mucosa. As shown in Figure 5, IκB protein levels were analyzed by Western blotting, respectively using cytoplasmic extracts of colonic mucosa from each model rat. 50% ethanol groups had a high level of cytoplasmic IκB (Lanes 1, 2). Intestinal inflammation of TNBS group induced signifficative IκB degradation (Lane 3). IκB degradations were both blocked by 0.5% SASP and 2% curcumin treatment (Lanes 4, 5, 6).

![Figure 3](image1.png) Results are shown as the intestinal histologic features of each group rats stained with HE. A: 50% ethanol group; B: TNBS group; C: 0.5 % SASP group; D: 2.0% curcumin preventive group; E: 2.0% curcumin therapeutic group.

**Effects of curcumin on expression of cytokine genes in the colonic mucosa in TNBS-induced colitis**

By reverse-transcription polymerase chain reaction analysis, our results showed that mRNA levels of proinflammatory cytokine, such as IL-1β, in colon specimens from rats with TNBS-induced colitis, was significantly higher than that in normal controls. Anti-inflammatory cytokine IL-10 level in colon specimens from rats with TNBS-induced colitis was relatively lower than that in normal control, but there was no significant difference between the two groups. High IL-1β mRNA expression was observably suppressed in colon specimens from curcumin-treated and SASP-treated rats with TNBS-induced colitis. Low IL-1β mRNA expression was observably increased in colon specimens from curcumin-treated and SASP-treated rats with TNBS-induced colitis (Figure 6).

![Figure 4](image2.png) Effects of curcumin on NF-κB activity of colonic mucosa in rats with TNBS-induced colitis. Lane 1: 50% ethanol group; lane 2: cold probe control group; lanes 3 and 4: TNBS group; lane 5: 0.5 % SASP group; lane 6: 2.0% curcumin preventive group; lane 7: 2.0% curcumin therapeutic group.

**DISCUSSION**

Homeostasis of TH1/TH2 type cytokines is important in intestinal mucosal immunity. T helper cells excrete a lot of cytokines and chemokines in normal intestinal tissue. They induce other types of cell infiltration, such as leukocytes, monocytes, macrophages and other lymphocytes. They induce inflammatory and immune responses aiming nosogenesis outside cells which could straightly or indirectly cause damage of intestinal epithelial cells. Most notable CD and UC and other chronic evolutional intestinal inflammation are all mediated by proinflammatory immune responses induced by diversified antigens. However, studies had made sure that TH1 type cytokines play a predominant role in intestinal mucosal immunity either in various animal models of IBD or in patients with IBD. Proinflammatory cytokines were significantly high expression and anti-inflammatory...
cytokines were low expression. Expression of proinflammatory cytokines in TH1 type immune response such as IL-1β, IL-6, IL-12, IL-18, TNF-α and IFN-γ in intestinal mucosa of animal models and humans IBD were significantly enhanced[10-13]. Expression of anti-inflammatory cytokines such as IL-4, IL-5 and IL-10 were low level[12-15].

In our animal experimental model of IBD induced by TNBS, treatment with 2% curcumin and 0.5% SASP can suppress proinflammatory cytokine IL-1β mRNA high expression and increase anti-inflammatory cytokine IL-10 mRNA low expression in colonic mucosa.

Canonical pathway of NF-κB activation selectively regulates diversified inflammatory cytokines chemotactic cytokines and conglutinant cytokines[16,17]. The movement of activation NF-κB into the cellular nucleus is controlled by the targeted phosphorylation and subsequent degradation of IκB. Studies have elaborated several important and unexpected findings that explain mechanisms of NF-κB activation. In the nucleus, NF-κB dimers bind to target DNA elements and activate transcription of genes encoding proteins involved with immune or inflammation responses and with cell growth control[18].

Previous observations provide evidence for IBD that NF-κB activation is significantly elevated in UC and CD[19]. SASP, mesalamine, and corticosteroids have been used in different formulations for the treatment of patients with both acute and chronic IBD in clinic during the past decades, which all are NF-κB inhibitors[20,21-23]. They have sundry defects such as steroid dependence and steroid resistance, decreasing glucose tolerance, hypohepatia and pancreatitis. Now, more potent and selective treatment strategies with decreasing glucose tolerance, hypohepatia and pancreatitis.

Defects such as steroid dependence and steroid resistance, curvature and yellow color to curry[23]. Curcumin is used as a spice to give the specific flavor[23]. Curcumin is the major constituent of turmeric powder extracted from the rhizomes of the plant Curcuma longa Linn. Curcumin is used as a spice to give the specific flavor[23]. Scientists had understood curcumin mechanism of action in regulating the NF-κB pathogenesis[24]. Curcumin suppresses lipopolysaccharide-induced cyclooxygenase-2 expression by inhibiting activator protein 1 and nuclear factor κB bindings in BV2 microglial cells[25]. Curcumin inhibits dose-dependently and time-dependently neuroglial cell proliferation and growth[26].

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